



# UK Standards for Microbiology Investigations

## Identification of *Pasteurella* species and Morphologically Similar Bacteria



## Acknowledgments

UK Standards for Microbiology Investigations (SMIs) are developed under the auspices of Public Health England (PHE) working in partnership with the National Health Service (NHS), Public Health Wales and with the professional organisations whose logos are displayed below and listed on the website

<http://www.hpa.org.uk/SMI/Partnerships>. SMIs are developed, reviewed and revised by various working groups which are overseen by a steering committee (see <http://www.hpa.org.uk/SMI/WorkingGroups>).

The contributions of many individuals in clinical, specialist and reference laboratories who have provided information and comments during the development of this document are acknowledged. We are grateful to the Medical Editors for editing the medical content.

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UK Standards for Microbiology Investigations are produced in association with:



## Contents

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ACKNOWLEDGMENTS .....	2
AMENDMENT TABLE .....	4
UK STANDARDS FOR MICROBIOLOGY INVESTIGATIONS: SCOPE AND PURPOSE.....	5
SCOPE OF DOCUMENT .....	8
INTRODUCTION .....	8
TECHNICAL INFORMATION/LIMITATIONS.....	9
1 SAFETY CONSIDERATIONS .....	10
2 TARGET ORGANISMS.....	10
3 IDENTIFICATION.....	10
4 IDENTIFICATION OF <i>PASTEURELLA</i> SPECIES AND MORPHOLOGICALLY SIMILAR BACTERIA .....	12
5 REPORTING .....	13
6 REFERRALS.....	13
7 NOTIFICATION TO PHE OR EQUIVALENT IN THE DEVOLVED ADMINISTRATIONS .....	14
REFERENCES .....	15



NICE has accredited the process used by Public Health England to produce Standards for Microbiology Investigations. Accreditation is valid for 5 years from July 2011. More information on accreditation can be viewed at [www.nice.org.uk/accreditation](http://www.nice.org.uk/accreditation).

For full details on our accreditation visit: [www.nice.org.uk/accreditation](http://www.nice.org.uk/accreditation).

## Amendment Table

Each SMI method has an individual record of amendments. The current amendments are listed on this page. The amendment history is available from [standards@phe.gov.uk](mailto:standards@phe.gov.uk).

New or revised documents should be controlled within the laboratory in accordance with the local quality management system.

Amendment No/Date.	8/11.03.14
Issue no. discarded.	2.2
Insert Issue no.	2.3
<b>Section(s) involved</b>	<b>Amendment</b>
Whole document.	<p>Document has been transferred to a new template to reflect the Health Protection Agency's transition to Public Health England.</p> <p>Front page has been redesigned.</p> <p>Status page has been renamed as Scope and Purpose and updated as appropriate.</p> <p>Professional body logos have been reviewed and updated.</p> <p>Standard safety and notification references have been reviewed and updated.</p> <p>Scientific content remains unchanged.</p>

Amendment No/Date.	7/21.10.11
Issue no. discarded.	2.1
Insert Issue no.	2.2
<b>Section(s) involved</b>	<b>Amendment</b>
Whole document.	Document presented in a new format.
References.	Some references updated.

## UK Standards for Microbiology Investigations<sup>#</sup>: Scope and Purpose

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### Users of SMIs

- SMIs are primarily intended as a general resource for practising professionals operating in the field of laboratory medicine and infection specialties in the UK.
- SMIs provide clinicians with information about the available test repertoire and the standard of laboratory services they should expect for the investigation of infection in their patients, as well as providing information that aids the electronic ordering of appropriate tests.
- SMIs provide commissioners of healthcare services with the appropriateness and standard of microbiology investigations they should be seeking as part of the clinical and public health care package for their population.

### Background to SMIs

SMIs comprise a collection of recommended algorithms and procedures covering all stages of the investigative process in microbiology from the pre-analytical (clinical syndrome) stage to the analytical (laboratory testing) and post analytical (result interpretation and reporting) stages.

Syndromic algorithms are supported by more detailed documents containing advice on the investigation of specific diseases and infections. Guidance notes cover the clinical background, differential diagnosis, and appropriate investigation of particular clinical conditions. Quality guidance notes describe laboratory processes which underpin quality, for example assay validation.

Standardisation of the diagnostic process through the application of SMIs helps to assure the equivalence of investigation strategies in different laboratories across the UK and is essential for public health surveillance, research and development activities.

### Equal Partnership Working

SMIs are developed in equal partnership with PHE, NHS, Royal College of Pathologists and professional societies.

The list of nominating societies may be found at <http://www.npa.org.uk/SMI/Partnerships>. Inclusion of a logo in an SMI indicates participation of the society in equal partnership and support for the objectives and process of preparing SMIs. Nominees of professional societies are members of the Steering Committee and Working Groups which develop SMIs. The views of nominees cannot be rigorously representative of the members of their nominating organisations nor the corporate views of their organisations. Nominees act as a conduit for two way reporting and dialogue. Representative views are sought through the consultation process.

SMIs are developed, reviewed and updated through a wide consultation process.

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<sup>#</sup>Microbiology is used as a generic term to include the two GMC-recognised specialties of Medical Microbiology (which includes Bacteriology, Mycology and Parasitology) and Medical Virology.

## Quality Assurance

NICE has accredited the process used by the SMI Working Groups to produce SMIs. The accreditation is applicable to all guidance produced since October 2009. The process for the development of SMIs is certified to ISO 9001:2008.

SMIs represent a good standard of practice to which all clinical and public health microbiology laboratories in the UK are expected to work. SMIs are NICE accredited and represent neither minimum standards of practice nor the highest level of complex laboratory investigation possible. In using SMIs, laboratories should take account of local requirements and undertake additional investigations where appropriate. SMIs help laboratories to meet accreditation requirements by promoting high quality practices which are auditable. SMIs also provide a reference point for method development.

The performance of SMIs depends on competent staff and appropriate quality reagents and equipment. Laboratories should ensure that all commercial and in-house tests have been validated and shown to be fit for purpose. Laboratories should participate in external quality assessment schemes and undertake relevant internal quality control procedures.

## Patient and Public Involvement

The SMI Working Groups are committed to patient and public involvement in the development of SMIs. By involving the public, health professionals, scientists and voluntary organisations the resulting SMI will be robust and meet the needs of the user. An opportunity is given to members of the public to contribute to consultations through our open access website.

## Information Governance and Equality

PHE is a Caldicott compliant organisation. It seeks to take every possible precaution to prevent unauthorised disclosure of patient details and to ensure that patient-related records are kept under secure conditions.

The development of SMIs are subject to PHE Equality objectives [http://www.hpa.org.uk/webc/HPAwebFile/HPAweb\\_C/1317133470313](http://www.hpa.org.uk/webc/HPAwebFile/HPAweb_C/1317133470313). The SMI Working Groups are committed to achieving the equality objectives by effective consultation with members of the public, partners, stakeholders and specialist interest groups.

## Legal Statement

Whilst every care has been taken in the preparation of SMIs, PHE and any supporting organisation, shall, to the greatest extent possible under any applicable law, exclude liability for all losses, costs, claims, damages or expenses arising out of or connected with the use of an SMI or any information contained therein. If alterations are made to an SMI, it must be made clear where and by whom such changes have been made.

The evidence base and microbial taxonomy for the SMI is as complete as possible at the time of issue. Any omissions and new material will be considered at the next review. These standards can only be superseded by revisions of the standard, legislative action, or by NICE accredited guidance.

SMIs are Crown copyright which should be acknowledged where appropriate.

**Suggested Citation for this Document**

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UNDER REVIEW

## Scope of Document

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This SMI describes the procedure for the phenotypic identification of *Pasteurella* species, and distinguishes these from morphologically similar species.

This SMI should be used in conjunction with other SMIs.

## Introduction

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### Taxonomy

Currently some 15-20 species are included in the genus *Pasteurella*. Not all of these are true members. DNA-DNA hybridisation indicates that some of the species are more closely related to the genus *Actinobacillus*<sup>1</sup>.

*Pasteurella multocida* is the type species of the genus.

### Characteristics<sup>2</sup>

The *Pasteurella* species are spherical, ovoid or rod-shaped cells 0.5-1.0µm in diameter and 1.0-2.0µm in length. Cells are Gram negative and occur singly, or in pairs or short chains. Bipolar staining may be seen and capsules may be present. All species are non-motile, and are facultatively anaerobic.

*Pasteurella* species have both an oxidative and fermentative metabolism. The optimum growth temperature is 37°C. Glucose and other carbohydrates are catabolised with the production of acid but no gas. Most species are catalase positive and oxidase positive; nitrates are reduced to nitrites by almost all species.

Colonies of *Pasteurella* species are usually grey and viscous, with a strong mucinous odour. Rough, irregular colonies may also occur. Freshly isolated strains of *Pasteurella haemolytica* produce clear zones of β-haemolysis on blood agar—this organism is a cause of mastitis and septicæmia in some peridomestic animals, but very rarely infects humans.

*Pasteurella* and *Actinobacillus* species are so similar that no single phenotypic feature reliably distinguishes between the two genera. In clinical practice, however, an organism with characteristics corresponding to the genus *Pasteurella* is highly likely to be so if recovered from clinical specimens in association with a bite from a cat or dog.

The genus *Actinobacillus* now includes *Actinobacillus ureae*, formerly *Pasteurella ureae*. *A. ureae* is thought to be a commensal or occasionally an opportunist pathogen of human beings, and has principally been reported in connection with disease of the respiratory tract (eg cases of pneumonia, lung abscess). Occasionally, invasive infections (bacteraemia, meningitis) have also been reported.

As the name suggests, *A. ureae* is urease positive. Most species of *Pasteurella* are urease negative (including *P. multocida*). Thus, a *Pasteurella*-like organism, urease positive, recovered in association with human respiratory tract disease, is likely to be *A. ureae*.

Phenotypically, *Pasteurella* species may resemble *Haemophilus* species, but *Pasteurella* species will not regularly exhibit satellitism around colonies of *Staphylococcus* species, nor are they regularly auxotrophic for X or V factors; growth is not especially enhanced by use of chocolate blood agar.



## Principles of Identification

Colonies on blood agar are identified by colonial morphology, Gram stain, oxidase test and catalase production. Additional tests are needed for confirmation and/or isolates should be referred to the Reference Laboratory.

## Technical Information/Limitations

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N/A

UNDER REVIEW

## 1 Safety Considerations<sup>3-19</sup>

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Refer to current guidance on the safe handling of all organisms documented in this SMI.

Laboratory procedures that give rise to infectious aerosols must be conducted in a microbiological safety cabinet.

The above guidance should be supplemented with local COSHH and task specific risk assessments.

Compliance with postal and transport regulations is essential.

## 2 Target Organisms

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***Pasteurella* species reported to have caused human infection<sup>1</sup>**

*P. aerogenes*

*P. bettyae*

*P. canis*

*P. dagmatis*

*P. multocida* subspecies *gallicida*

*P. multocida* subspecies *multocida*

*P. multocida* subspecies *septica*

*P. pneumotropica*

*P. stomatis*

*P. trehalosi* (previously *P. haemolytica* biotype T)

*Avibacterium gallinarum* (formerly *P. gallinarum*)

*Mannheimia haemolytica* (formerly *P. haemolytica* (Biotype A))

## 3 Identification

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### 3.1 Microscopic Appearance

**Gram stain ([GP 29 - Staining Procedures](#))**

Spherical, ovoid or rod-shaped Gram negative cells which occur singly or in pairs or short chains. Bipolar staining is common, capsules may be present.

### 3.2 Primary Isolation Media

Blood agar 16–48hr incubation in 5-10% CO<sub>2</sub> at 35-37°C.

### 3.3 Colonial Appearance

Colonies are grey and viscous but rough irregular colonies occur frequently. Freshly isolated strains of *M. haemolytica* produce clear zones of β-haemolysis on blood agar.

### 3.4 Test Procedures

#### Oxidase test ([TP 26 - Oxidase Test](#))

Positive (almost always).

#### Catalase test ([TP 8 - Catalase Test](#))

Positive.

#### Growth on CLED or MacConkey

No growth (*P. multocida*) on MacConkey but can grow poorly on some CLED agar.

#### Sensitivity to penicillin

A zone of inhibition around a 1U penicillin disc may aid differentiation from other Gram negative bacilli.

#### Commercial identification kit

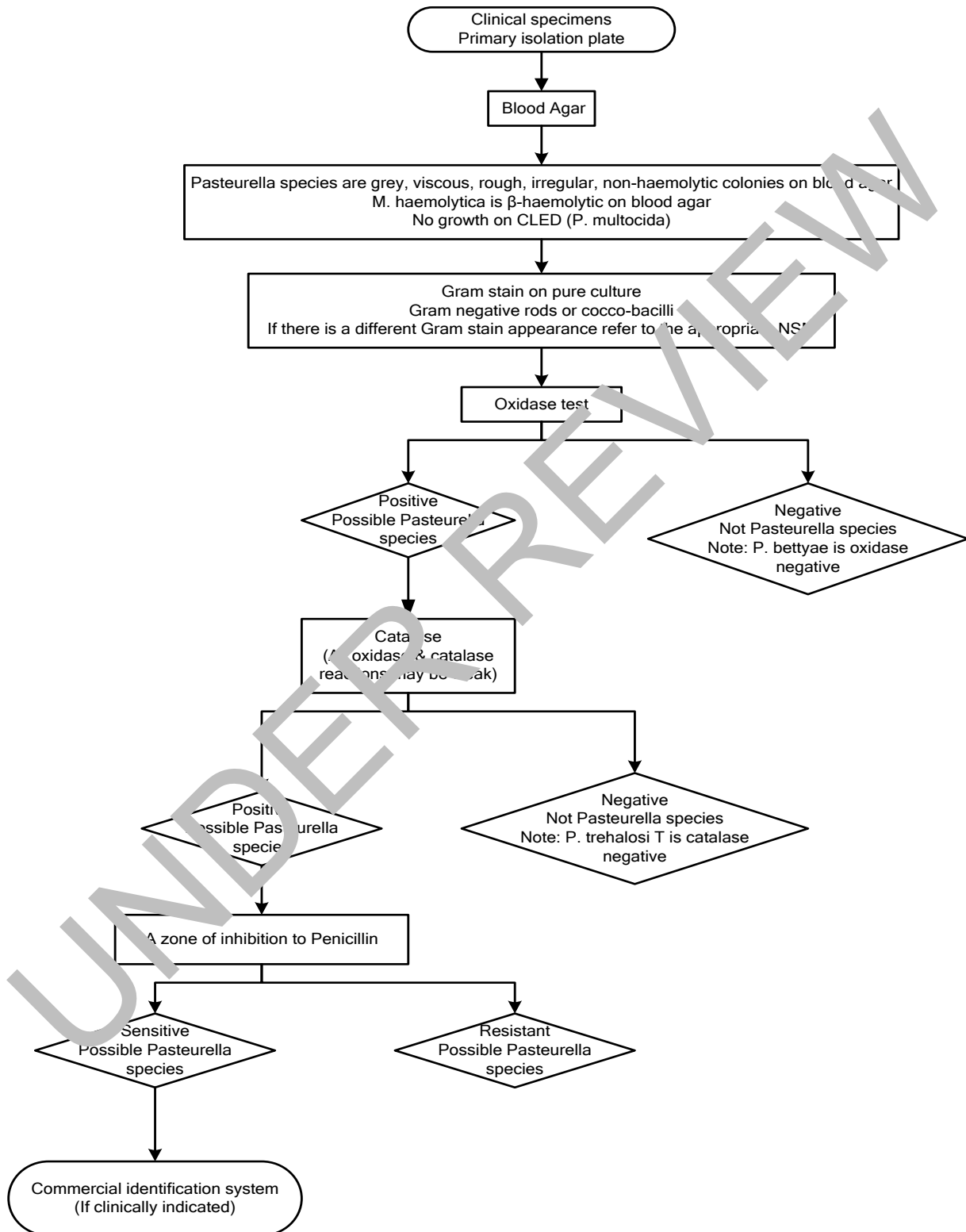
### 3.5 Further Identification

Following use of a commercial characterisation kit and/or referral to a Reference Laboratory.

### 3.6 Storage and Referral

If required save pure isolate on a blood agar slope for referral to the Reference Laboratory.

## 4 Identification of *Pasteurella* species and Morphologically Similar Bacteria



The flowchart is for guidance only

## 5 Reporting

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### 5.1 Presumptive Identification

If appropriate growth characteristics, colonial appearance and Gram stain of the culture are demonstrated.

### 5.2 Confirmation of Identification

N/A

### 5.3 Medical Microbiologist

The medical microbiologist should be informed of presumptive or confirmed *Pasteurella* species if isolated from a specimen from a normally sterile site or from other specimens in accordance with local protocols.

Follow local protocols for reporting to clinician.

### 5.4 CCDC

Refer to local Memorandum of Understanding.

### 5.5 Public Health England<sup>20</sup>

Refer to current guidelines on CDSC and GOC/JRV reporting.

### 5.6 Infection Control Team

N/A

## 6 Referrals

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### 6.1 Reference Laboratory

Contact appropriate devolved nation reference laboratory for information on the tests available, turn around times, transport procedure and any other requirements for sample submission:

Antimicrobial Resistance and Healthcare Associated Infections Reference Unit  
(AMRU)

Microbiology Services Division

Microbiology Services

Public Health England

61 Colindale Avenue

London

NW9 5EQ

<http://www.hpa.org.uk/cfi/lhcai/default.htm>

Contact PHE's main switchboard: Tel. +44 (0) 20 8200 4400

England and Wales

<http://www.hpa.org.uk/webw/HPAweb&Page&HPAwebAutoListName/Page/1158313434370?p=1158313434370>

Scotland

<http://www.hps.scot.nhs.uk/reflab/index.aspx>

Northern Ireland

<http://www.belfasttrust.hscni.net/Laboratory-MortuaryServices.htm>

## 7 Notification to PHE<sup>20,21</sup> or Equivalent in the Devolved Administrations<sup>22-25</sup>

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The Health Protection (Notification) regulations 2010 require diagnostic laboratories to notify Public Health England (PHE) when they identify the causative agents that are listed in Schedule 2 of the Regulations. Notifications must be provided in writing, on paper or electronically, within seven days. Urgent cases should be notified orally and as soon as possible, recommended within 24 hours. These should be followed up by written notification within seven days.

For the purposes of the Notification Regulations, the recipient of laboratory notifications is the local PHE Health Protection Team. If a case has already been notified by a registered medical practitioner, the diagnostic laboratory is still required to notify the case if they identify any evidence of an infection caused by a notifiable causative agent.

Notification under the Health Protection (Notification) Regulations 2010 does not replace voluntary reporting to PHE. The vast majority of NHS laboratories voluntarily report a wide range of laboratory diagnoses of causative agents to PHE and many PHE Health protection Teams have agreements with local laboratories for urgent reporting of some infections. This should continue.

**Note:** The Health Protection Legislation Guidance (2010) includes reporting of Human Immunodeficiency Virus (HIV) & Sexually Transmitted Infections (STIs), Healthcare Associated Infections (HCAIs), and Creutzfeldt–Jakob disease (CJD) under 'Notification Duties of Registered Medical Practitioners': it is not noted under 'Notification Duties of Diagnostic Laboratories'.

Other arrangements exist in Scotland<sup>22,23</sup>, Wales<sup>24</sup> and Northern Ireland<sup>25</sup>.

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## Identification of *Pasteurella* species and Morphologically Similar Bacteria

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